Saccade Dysmetria during Functional Perturbation of the Caudal Fastigial Nucleus in the Monkey

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ABSTRACT: The caudal fastigial nucleus (cFN) is the output nucleus by which the medioposterior cerebellum influences the brainstem saccade generator. In the monkey, inactivation of one cFN by local injection of muscimol impairs all saccades: ipsiversive saccades become hypermetric, contraversive saccades become hypometric, and saccades aimed at a target located in the upper or lower visual fields are biased horizontally toward the injected side. The pharmacological action of muscimol does not allow deficits that are presaccadic to be distinguished from those occurring during saccade execution. To determine the interval during which altered cFN activity affects saccade accuracy, we applied low-frequency electrical microstimulation (100 Hz for 100–300 ms) to the cFN of three monkeys while they were making saccades toward a flashed target. Similar to the effect of muscimol injection in cFN, low-frequency microstimulation biased all saccades toward the ipsilateral side. When the microstimulation was applied after target flash and before saccade onset, the ipsilateral bias was absent. However, when the stimulation was applied during the ongoing movement, the saccade trajectory was biased toward the stimulated side. The muscimol-like effect of the microstimulation suggests that the stimulation inhibits cFN activity, possibly by recruiting the inhibitory afferents from the cerebellar vermis (axons of Purkinje cells). Low-frequency microstimulation had to be applied during the saccade to bias its trajectory. These data suggest that the ipsilateral horizontal bias observed during muscimol inactivation results from an imbalance in the intrasaccadic activity between the two caudal fastigial nuclei.

KEYWORDS: saccades; cerebellum; fastigial; reversible inactivation; microstimulation; monkey

INTRODUCTION

The medioposterior cerebellum is one of the most important brain regions involved in the transformation of target-related visual signals into motor commands that move the line of sight accurately toward the target location. Indeed, damage of this cerebellar region severely impairs the accuracy of goal-directed gaze shifts (for
In the head-restrained monkey, the unilateral disinhibition of neuronal activity in the caudal fastigial nucleus (cFN) by local injection of bicuculline or by lesion of the vermal lobules VI–VII impairs the accuracy of visually triggered saccades. Saccades toward the disinhabited side (ipsiversive saccades) become hypometric, and saccades toward the opposite side (contraversive saccades) become hypermetric. Conversely, the inactivation of the cFN by local injection of muscimol leads to hypermetric ipsiversive saccades and hypometric contraversive saccades. In the head-unrestrained cat, after unilateral injection of muscimol in the cFN, ipsiversive combined eye-head gaze shifts become hypermetric, and contraversive ones become hypometric, without change in the contribution of the head to the overall gaze displacement.

Although most cFN neurons show a steady firing rate during intersaccadic intervals, the dysmetria observed during muscimol injection in the cFN has been proposed to result from the suppression of saccade-related bursts of activity in cFN. According to this hypothesis, the hypermetria of ipsiversive saccades would be due to the suppression of the burst generated during the late period of the saccades, whereas the hypometria of contraversive saccades would be due to the suppression of the burst that precedes the onset of saccades.

An alternative hypothesis, based on several observations made in the head-unrestrained cat, has been proposed to account for the dysmetria resulting from unilateral muscimol injections into cFN. According to this hypothesis, the dysmetria results from an impaired specification of the movement metrics prior to movement onset, rather than an intrasaccadic deficit. Indeed, the observation of misdirected and inappropriately initiated ipsiversive gaze shifts suggests more than a deficit in the control of movement deceleration. All these ipsiversive gaze shifts overshoot the target, producing a constant horizontal error: the endpoints are horizontally shifted with respect to the target position, irrespective of the starting gaze position. Moreover, no consistent modifications in the dynamics of the eye, head, and gaze displacements or in the eye-head coupling are observed. Occasional muscimol-induced modifications in dynamics are unrelated to the magnitude of the dysmetria. Finally, changes in the latency of the gaze and head displacements during cFN inactivation or after ablation in the oculomotor vermis support the idea of a deficit unfolding prior to movement onset.

Some observations in the head-restrained monkey suggest that muscimol injection in the cFN can lead to deficits in the primate that are similar to those observed in the cat. In particular, saccades initiated from various eccentric eye positions and aimed at a visual target located straight ahead all end at approximately the same final position, one that is horizontally shifted relative to the target. This is very reminiscent of the constant horizontal error observed in the cat (see Figure 2H in the article by Ohtsuka and colleagues).

PERTURBING THE CAUDAL FASTIGIAL NUCLEUS BY LOCAL INJECTION OF MUSCIMOL

The effects of inactivating the saccade-related area in the cFN was studied in three head-restrained monkeys to determine if saccades initiated from various start-
FIGURE 1. Effects on saccade direction and amplitude of perturbation of the cFN by local injection of muscimol. The saccades were aimed at flashed (duration = 100 ms) target LEDs (circles) located (A) 16°, (B) 12°, or (C) 6° on the vertical meridian. Variation in the starting position of the eyes was obtained by using seven different fixation LEDs (diamonds): ±12°, ±8°, ±4°, and 0° along the horizontal meridian. For clarity, saccades starting from fixation LEDs located to the left are plotted with a different symbol (open) from those starting from fixation LEDs located to the right (filled). Volume injected: 0.8 µL (1 µg/µL).
ing positions and aimed at the same visual target missed the target with a similar constant horizontal error. Figure 1 shows the effect of injecting a small volume of muscimol in the cFN on the trajectories of representative saccades initiated from various starting positions along the horizontal meridian and aimed at a brief target light-emitting diode (LED) (duration = 100 ms). Target LEDs (circles) were located 16° (A), 12° (B), or 6° (C) on the vertical meridian (Fig. 1A–C, respectively). The left panel shows the fixation and the saccade performance before the injection (control). After muscimol injection in the left cFN (right panel), a leftward offset in starting eye position was observed when the animal was viewing the fixation LEDs. This fixation offset was to the right following the injection in the right cFN (results not shown, but see Robinson and colleagues5 and Goffart and Pélisson9). With respect to saccades toward the 16° upward target (Fig. 1A), the horizontal component of leftward (ipsiversive) saccades was too large, whereas the horizontal component of rightward (contraversive) saccades was too short to acquire the target. Moreover, after the muscimol injection, the initial direction of saccades starting from the straight-ahead fixation LED was to the left, and saccades in this direction increased, rather than decreased, the horizontal distance between gaze and target positions. Saccades initiated from the 4° leftward fixation LED had no rightward component even though the target for these trials was located more than 4° to the right. Saccades starting from fixation LEDs located further to the left had less of a rightward initial direction than would be required for target acquisition. The amplitude of their horizontal component was hypometric. The initial direction of saccades starting from fixation LEDs located to the right had more of a leftward component than that required to look to the target. The amplitude of the horizontal component was hypermetric with an overshoot that increased with more eccentric starting position (or with larger horizontal target eccentricity). Similar effects upon the initial direction were apparent for saccades directed to the 12° and 6° targets (Fig. 1B, C, respectively). However, when the vertical eccentricity of the target was reduced, the horizontal error at the end of the saccades was also significantly reduced (−3.9 ± 0.7, −2.9 ± 0.7, and −1.9 ± 0.5° for targets 16°, 12°, and 6° upward, respectively; Mann–Whitney U test, P < 0.01 for each comparison). The endpoints of saccades starting from the straight-ahead fixation LED were offset relative to the target with a magnitude that increased with the vertical target eccentricity (see also Iwamoto7). Moreover, when the saccades initiated from the most eccentric fixation LEDs to the left (−8° or −12°) were examined, the amplitudes of their horizontal components were larger when the vertical target position decreased from 16° to 6° upward.

In summary, saccades initiated from various starting positions did not end at the same final eye position that was shifted relative to the target location by a constant horizontal error. Rather, ipsiversive saccades missed the target with a horizontal error that increased as the eccentric starting deviation of the eyes in the orbit increased and, concomitantly, as horizontal target eccentricity and saccade duration increased. Statistical analysis indicates that the horizontal error of oblique ipsiversive saccades aimed at target with similar eccentricity (for example, 12° leftward and upward) was not statistically different between saccades initiated from the 12° eccentric fixation LED and saccades initiated from the central fixation LED (Mann–Whitney U test, P level >0.10). Contraversive saccades toward one given target seemed to reach the same final eye position, but the location of saccade endpoints was different for different targets.
From the inactivation data, it is impossible to determine whether the dysmetria is due to presaccadic or intrasaccadic imbalances in the activity of the two cFNs. Thus, we developed a technique that allowed more transient inhibition of the activity of one cFN. This technique relies on the assumption that electrical microstimulation of the dorsocaudal portion of the fastigial nucleus inhibits cFN neurons by activating the inhibitory axons of Purkinje cells from the vermis.15 By manipulating the duration of the microstimulation and its delay with respect to the target presentation, we were able to determine the critical period during which cFN activity influences saccade accuracy within the time interval between the onset of the target and the end of the saccade.

FIGURE 2 illustrates the effect of microstimulating the dorsocaudal portion of the fastigial nucleus. The stimulation train (black horizontal bar; 100-Hz, 0.2-ms pulse; duration = 100 ms in A, 200 ms in B) was applied while the animal was waiting for the appearance of a visual target. Current intensity was 35 µA and 20 µA for the left cFN and right cFN of the same animal, respectively.

PERTURBING THE CAUDAL FASTIGIAL NUCLEUS BY LOW-FREQUENCY ELECTRICAL MICROSTIMULATION

From the inactivation data, it is impossible to determine whether the dysmetria is due to presaccadic or intrasaccadic imbalances in the activity of the two cFNs. Thus, we developed a technique that allowed more transient inhibition of the activity of one cFN. This technique relies on the assumption that electrical microstimulation of the dorsocaudal portion of the fastigial nucleus inhibits cFN neurons by activating the inhibitory axons of Purkinje cells from the vermis.15 By manipulating the duration of the microstimulation and its delay with respect to the target presentation, we were able to determine the critical period during which cFN activity influences saccade accuracy within the time interval between the onset of the target and the end of the saccade.

FIGURE 2 illustrates the effect of microstimulating the dorsocaudal portion of the left and right fastigial nucleus with a 100-Hz train during a gap interval (500 ms) when the animal was waiting for the appearance of a visual target. Eye position was stable during the entire period of microstimulation (black horizontal bar; duration = 100 ms in Fig. 2A, 200 ms in Fig. 2B), but a “rebound” saccade was generated 40 to 60 ms after the offset of the microstimulation train. This rebound saccade was always directed toward the side contralateral to the stimulated side, that is, rightward.
When the left cFN was stimulated (left panels in Fig. 2), and leftward when the right cFN was stimulated (right panels in Fig. 2). Previous work has shown that contralateral saccades are also evoked by microstimulating the axons of cFN neurons. The contralateral saccades illustrated in Figure 2 could thus result from the rebound depolarization and spike bursting by cFN neurons following a prolonged microstimulation-induced hyperpolarization. When applied while the monkey was preparing a goal-directed saccade, the microstimulation train had an effect on saccade trajectory that did indeed suggest that the microstimulation inhibits the activity in the cFN.

FIGURE 3 shows the effect of a microstimulation train at 100 Hz on the trajectory of saccades initiated from different horizontal starting positions and aimed at a brief target (duration = 100 ms) presented 16° above or below the central fixation LED. The stimulation train was synchronized with respect to the target offset. It overlapped the interval during which the line of sight was moving toward the target. Variations in the horizontal starting position of the eyes were obtained by using five different fixation LEDs (diamonds): ±10°, ±5°, and 0° along the horizontal meridian. Stimulation parameters for left cFN (same site as in Fig. 2, left panels): 100 Hz, 100 or 200 ms, 20 µA; stimulation parameters for right cFN: 100 Hz, 300 ms, 100 µA.

FIGURE 4 shows the effect of varying the interval during which the microstimulation was applied upon the trajectory and the accuracy of saccades. In the trials
FIGURE 4. Effects of varying the period of stimulation within the target-offset/saccade-end interval. The fixation LED was located straight ahead, and the target LED was flashed 16° above the fixation stimulus. The microstimulation (black horizontal bar; 100-Hz, 0.2-ms pulse; 100-ms duration; 45 μA) was applied (A) immediately, (B) 50 ms, or (C) or 100 ms after the target LED was extinguished. The x-y plots of eye position are shown on the left, and plots of horizontal (H) and vertical (V) eye positions as a function of time are shown on the right. The black horizontal bar represents the period when microstimulation was delivered. The traces are aligned on the onset of the microstimulation train.
shown as examples, the fixation LED was located straight ahead, and the target LED was flashed 16° above the fixation LED. The microstimulation was applied immediately, 50 ms, or 100 ms (Fig. 4A–C, respectively) after the target LED was extinguished. The stimulation duration was set to 100 ms so that on some trials it could be restricted to the period prior to saccade initiation. In that particular case (Fig. 4A), only small effects on vertical saccades were observed: a slight final deviation away from the stimulation side occurred. When the microstimulation was applied during the overall saccade period (Fig. 4B), a much larger leftward bias was observed in the trajectory. A rightward rebound saccade immediately followed the stimulation offset. The ipsilateral bias in saccade trajectory was less important when the microstimulation was applied slightly before saccade onset (Fig. 4C). The figure also shows that the amount of microstimulation (duration) applied before (up to 10 ms before) the saccade was launched toward the visual target is not critical in determining the amount of deviation in saccade trajectory.

CONCLUSION

Low-frequency microstimulation of the dorsocaudal portion of the fastigial nucleus biases the trajectory of saccades toward the stimulated side. This effect is similar to the effect of muscimol injection in the cFN. The microstimulation must be applied during the saccade to bias its trajectory (see also Keller and colleagues17). More experiments using a larger range of intervals between stimulation onset and saccade onset are needed. But when an efferent delay of approximately 10 ms is taken into account, our data suggest that it is only the microstimulation immediately before saccade onset and during the execution period that is effective in modifying the trajectory of a saccade.

Further experiments are required to verify that these conclusions also hold in the head-unrestrained monkey. If so, the particular dysmetria observed in the head-unrestrained cat9 may be due to the nature of the behavioral responses evoked by the target. Indeed, in the cat experiments, the food target was triggering an orienting gaze shift but also an orienting movement of the mouth (and thus of the head). In the monkey experiments, target acquisition required only a shift of the line of sight.

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